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# Pattern of Fungal Colonization and Co-occurrence on *Avicennia officinalis* Woody Litter in a Southwestern Mangrove of India

G.L. Maria and K.R. Sridhar\*

Department of Botany, St. Agnes College, Mangalore 575 002, Karnataka, India Department of Biosciences, Mangalore University, Mangalagangotri 574 199, Mangalore, Karnataka, India \*Corresponding author Email: kandikere@gmail.com (Submitted in January, 2017; Accepted on June 17, 2017)

#### ABSTRACT

This study outlines the pattern of colonization and co-occurrence of saprophytic fungi on woody litter of *Avicennia officinalis* tree species in Nethravathi mangrove. Two types of woody litter (naturally and artificially submerged) during two seasons (monsoon and summer) were assessed. Natural woody litter consists of 57 fungal species, while it was 34 species in submerged woody litter. Composition of fungi was a mosaic of typical marine and terrestrial fungi. Higher number of fungi (teleomorphs, anamorphs and core-group species) was found in natural than submerged wood, so also during monsoon than summer. Eight species (*Aniptodera* sp., *Lignincola laevis, Lulworthia grandispora, Savoryella lignicola, S. paucispora, Tricladium linderi, Tirispora* sp. and *Zalerion varia*) were common to natural and submerged wood in both seasons. The Shannon diversity was higher during monsoon than summer irrespective of type of wood assessed. The co-occurrence of fungi per wood was maximum up to 8 and 11 species during monsoon in submerged and natural wood, respectively. The pattern of three fungal species co-occurrence was also similar to overall colonization on woody litter. In both wood types, three species co-occurrence was the highest in both seasons. Per cent co-occurrence of core-group species showed dominance in natural than in submerged wood, so also during monsoon than summer. Single species occurrence of core-group fungi on woody litter was as low as 0% (16 core-group fungi on either of wood or season) and 12% (*Lignincola laevis*). The core-group fungus *L. laevis* co-occurrence up to a maximum of 72% during summer in natural wood which reveals its compatibility and accommodating ability with other associated fungi. Overall, this study reveals that the colonization and co-occurrence of fungi season.

Keywords: Diversity, mangrove fungi, species assemblage, species consortia, core-group fungi, monsoon, summer

#### **INTRODUCTION**

Mutualism is one of the evolutionary consequences which facilitated biological entities to live harmoniously to face unusual ecological and abiotic consequences. Mutualistic association of two or more species in an ecological niche is very common (e.g. plant species vs. arbuscular mycorrhizae/ectomycorrhizae/endophytic fungi). Among saprophytic fungi, co-occurrence or succession is inevitable to exploit detritus to a maximum extent. In this context, mangrove ecosystems provide excellent platform with wide variation in ecological, biotic and abiotic factors to investigate the activities and functions of saprophytic fungi (assemblage, diversity and community structure) (Cooke and Rayner, 1984). Decomposer fungal community in mangroves plays a major role in unlocking energy from the detritus and transfers it into the higher trophic levels (Wafar et al., 1997). Woody litter is one of the long-lasting resources in mangroves and provides niches for saprophytic fungi and fauna for sustained activity for extended periods. Mangrove woody litter constitutes the second important host after marine driftwood for fungal colonization (Hyde et al., 2000). Assessment of structural (e.g. assemblage, diversity and cooccurrence) and functional (e.g. decomposition, extracellular enzymes and respiration) profiles will be immensely valuable in assessment of ecosystem functions or fungal saprophytism. Although several studies are available on the diversity and activity of saprophytic fungi on woody litter in mangroves of east coast and west coast of India (e.g. Borse, 1988; Sarma and Vittal, 2000, 2001; Maria and Sridhar, 2003, 2004; Ananda and Sridhar, 2004), a few studies have addressed the co-occurrence (e.g. Sridhar and Maria, 2006; Sarma and Raghukumar, 2013). Thus, the main aim of this study is to document fungal assemblage and co-occurrence on woody litter (natural and introduced) of one of the dominant mangrove tree species Avicennia officinalis during two

seasons (monsoon and summer) in a southwestern mangrove.

### **MATERIALS AND METHODS**

Woody litter: Avicennia officinalis (Acanthaceae) is one of the dominant tree species in Nethravathi mangroves of the southwest coast of India (12°50'N, 74°50'E). Nethravathi mangrove show humid climate during June-November (monsoon) and dry climate during December-May. Naturally deposited trapped 150 woody litters of A. officinalis were sampled during wet season (October, 2014) and dry season (April, 2015). Easily breakable dried twigs of A. officinalis of uniform diameter (1.5 cm) were collected during February 2015, cut into segments (15 cm), sun-dried and preserved in polythene bags until submersion. In each nylon mesh bags (25 10 cm; mesh size, 1 mm), five wood pieces were inserted and 25 bags were randomly introduced in different locations by tying to roots at mid-tide level on the onset of monsoon up to four months (June-September, 2014) and in summer for four months (December, 2014-March, 2015). The introduced woody litters were sampled during wet and dry seasons (October, 2014 and April 2015, respectively). Similar to introduced wood, naturally submerged wood were collected during monsoon and summer were cut into desired size prior to incubation. Within eight hr of sampling, randomly assorted 100 each of natural and introduced woody litter were incubated (25±2°C) individually on sterile sand bath consisting dilute seawater (17‰) in sterile polythene bags. The incubated woody litters were screened for occurrence of fungal structures at fortnightly intervals up to four months. On each observation, the sand bath was rewetted with sterile distilled water. The fungi observed were identified using taxonomic descriptions (e.g. Hyde and Sarma, 2000; Sarma and Hyde, 2000; Jones et al., 2009).

Data analysis: Colonization frequency (CF%) of each fungus on natural and submerged woody litter during

monsoon and summer seasons were assessed [CF% = (Number of wood colonized by a specific fungus/Total number of wood supporting sporulating fungi) 100]. Shannon's diversity (Magurran, 1988) and Pielou's equitability (Pielou, 1975) of fungi colonized on woody litter were also assessed.

# RESULTS

**Colonization and diversity:** Natural woody litter consists of 57 species encompassing 31 teleomorphs and 26 anamorphs (**Table 1**). Higher number of fungi (teleomorphs and anamorphs) was found during monsoon (47 spp.) than summer season (21 spp.). Among the core-group fungi (>10%), 12 species were found in monsoon and seven species in summer (3 were core-group in both seasons).

Submerged wood consists of 34 species encompassing 19 teleomorphs and 15 anamorphs (**Table 1**). Monsoon sampled submerged wood consists of higher fungi (17 spp.) than summer (13 spp.). Eight core-group fungi were found in monsoon, while seven fungi in summer (6 were core-group in both seasons). Eight species (*Aniptodera* sp., *Lignincola laevis, Lulworthia grandispora, Savoryella lignicola, S. paucispora, Tirispora* sp., *Tricladium linderi* and *Zalerion* 

 Table 1. Colonization frequency (%) of fungi on Avicennia officinalis woody litter (-, not occurred; \*, core-group fungi at least on any woody litter or season).

| laxon   | Natural wood   |   | Submerged wood  |  |  |
|---|--|---|---|--|--|
|   | Monsoon  | Summer  | Monsoon   | Summer   |  |
| Ascomycetes   |  |   |   |  |  |
| *Aigialus mangroveis Borse  | -  | -   | 6   | 13   |  |
| Aniptodaya ahasanaakansis Shanyar & M.A. Mill   | 1  |   | 0   | 15   |  |
| A manarovei K D Hyde  | 2  | 2   | 2   | -  |  |
| A. mangrover K.D. riyac   | 2  | 2   | 4   | -  |  |
| Aniptodera sp   | 1  | 2   | 8   | 6  |  |
| Caryosporella rhizophorae Kohlm.  | 1  | -   | -   | -  |  |
| C. maritima Werderm.  | -  | -   | -   | 1  |  |
| Cumulospora sp.   | -  | -   | 1   | -  |  |
| Dactylospora haliotrepha (Kohlm. & E. Kohlm.) Hafellner   | -  | 1   | -   | -  |  |
| Didymella avicenniae S.D. Patil & Borse   | -  | 1   | -   | -  |  |
| Durella sp.   | 1  | -   | -   | -  |  |
| Eutypa bathurstensis K.D. Hyde & Rappaz   | 1  | -   | -   | -  |  |
| Halosarpheia cincinnatula Shearer & J.L. Crane  | -  | 1   | 4   | 7  |  |
| H. fibrosa Kohlm, & E. Kohlm.   | 2  | -   | -   | -  |  |
| H fibrosa Kohlm & E Kohlm   | 7  | -   | -   | _  |  |
| Kallichroma tethys (Kohlm) Kohlm & Volkm Kohlm  | 2  | -   | -   | _  |  |
| Lentosphaeria australiensis (Cribb & LW, Cribb) G.C. Hughes   |  | 2   | _   | -  |  |
| * <i>Lignineola lagyis</i> Höhnk  | 17   | 84  | 28  | 22   |  |
| Lagrandona intervisionink   | 17   | 04  | 20  | 22   |  |
| L. longirosiris (Chob & J. W. Chob) Kohini.   | 11   | -   | 4   | -  |  |
| L. tropica Konim.   |  | -   | 5   | -  |  |
| "Luiworinia grandispora Meyers  | 7  | 37  | 3   | 2  |  |
| *Lulworthia sp.   | -  | 8   | 39  | 49   |  |
| *Passeriniella mangrovei G.L. Maria & K.R. Sridhar  | 42   | -   | 8   | 3  |  |
| Saagaromyces ratnagiriensis (S.D. patil & Borse) K.L. Pang &  | 3  | -   | -   | -  |  |
| E.B.G. Jones  |  |   |   |  |  |
| *Savorvella lignicola E.B.G. Jones & R.A. Eaton   | 19   | 10  | 5   | 4  |  |
| S. longisporg E.B.G. Jones & K.D. Hyde  | 3  | 3   |   | -  |  |
| *S paucispora (Cribb & LW Cribb) L Koch   | 28   | 26  | 17  | 3  |  |
| Showsching an   | 20   | 20  | 1   | 2  |  |
| Twispore mandoviana VV Sarma & K.D. Uuda  | -  | -   | -   | 2  |  |
| *This and a second with a v.v. Sama & K.D. Hyde   | -  | 22  | 22  | 20   |  |
| * <i>Thrispora</i> sp.  | 4  | 23  | 33  | 38   |  |
| "Verrucuina enalia (Konim.) Konim. & Voikm. Konim.  | 4  | -   | 14  | 17   |  |
| Zignoëlla sp.   | 1  | -   |   | -  |  |
| Zopfiella latipes (N. Lundq.) Malloch & Cain  | 1  | -   | 1   | -  |  |
| *Ascomycete sp. 1   | 29   | -   | -   | -  |  |
| Ascomycete sp. 2  | 2  | -   | -   | -  |  |
| Ascomycete sp. 3  | 1  | -   | -   | -  |  |
| Ascomycete sp. 4  | -  | 8   | -   | -  |  |
| Basidiomycete   |  |   |   |  |  |
| Auricularia sp.   | 1  | -   | -   | -  |  |
| Anamorphs   |  |   |   |  |  |
| Acremonium sp.  | 3  | -   | 6   | -  |  |
| Alternaria sp. 1  | 3  | -   | 6   | 1  |  |
| Alternatia sp. 1  | 1  |   | 0   | 1  |  |
| Anernaria sp. 2   | 1  | -   | -   | -  |  |
| Brachysporiella gayana Bat.   | 1  | -   | -   | -  |  |
| Cirrenalia macrocephala (Kohlm.) Meyers & R.T. Moore  | 1  | -   | -   |  |  |
| *Cirrenalia pygmea Kohlm.   | _  |   |   |  |  |
| *C. tropicalis Kohlm.   |  | 27  | 15  | 22   |  |
|   | -  | 27<br>6   | 15<br>19  | 22   |  |
| Dactylella sp.  | 1  | 27<br>6<br>-  | 15<br>19  | 15   |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad  | -<br>1<br>3  | 27<br>6<br>-  | 15<br>19  | 22<br>15<br>-  |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.  | -<br>1<br>3<br>3   | 27<br>6<br>-<br>-   | 15<br>19<br>-   | 22<br>15<br>-  |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomvces roseus Link   | 1<br>3<br>3  | 27<br>6<br>-<br>-   | 15<br>19<br>-<br>-  | 22<br>15<br>-<br>-   |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Mondictws mutredinis (Wallr.) S. Hughes   | 1<br>3<br>3<br>1   | 27<br>6<br>-<br>-<br>-  | 15<br>19<br>-<br>-<br>-<br>2  | 22   |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes  | 1<br>3<br>3<br>1<br>10   | 27<br>6<br>-<br>-<br>-  | 15<br>19<br>-<br>-<br>2   | 22<br>15<br>-<br>-<br>-  |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys sp.  | 1<br>3<br>1<br>10<br>-   | 27<br>6<br>-<br>-<br>-<br>-   | 15<br>19<br>-<br>-<br>2<br>1  | 22<br>15<br>-<br>-<br>-<br>-   |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Penicillium sp.   | -<br>1<br>3<br>1<br>10<br>-<br>2   | 27<br>6<br>-<br>-<br>-<br>-   | 15<br>19<br>-<br>-<br>2<br>1<br>2   | 22<br>15<br>-<br>-<br>-<br>-   |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Penicillium sp.<br>Pericoila prolifica Anastasiou   | 1<br>3<br>1<br>10<br>-<br>2<br>1   | 27<br>6<br>-<br>-<br>-<br>-   | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2  | 22<br>15<br>-<br>-<br>-<br>-<br>1  |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Pericillum offica Anastasiou<br>*Phaeoistria clematidis (Fuckel) S. Hughes  | -<br>1<br>3<br>1<br>10<br>-<br>2<br>1<br>17  | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-   | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2  | 22<br>15<br>-<br>-<br>-<br>1   |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Penicillium sp.<br>Pericoila prolifica Anastasiou<br>*Phaeoisaria clematidis (Fuckel) S. Hughes<br>*Phome sp.   | 1<br>3<br>1<br>10<br>-<br>2<br>1<br>17<br>24   | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>-   | 22<br>15<br>-<br>-<br>-<br>1<br>-  |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Periciolia prolifica Anastasiou<br>*Phaeoisaria clematidis (Fuckel) S. Hughes<br>*Phaeoas.<br>Sporidesmium sp.  | 1<br>3<br>1<br>10<br>-<br>2<br>1<br>17<br>24<br>1  | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-                               | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>-   | 22<br>15<br>-<br>-<br>-<br>1<br>-  |  |
| Dactylella sp.<br>Diplociadiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Peniciallium sp.<br>Pericoila prolifica Anastasiou<br>*Phaeoisaria clematidis (Fuckel) S. Hughes<br>*Phoma sp.<br>Sporidesmium sp.  | -<br>1<br>3<br>3<br>1<br>10<br>-<br>2<br>1<br>17<br>24<br>1<br>12  | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>-<br>-<br>-<br>2<br>0   | 22<br>15<br>-<br>-<br>-<br>1<br>-<br>-<br>6  |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Penicillium sp.<br>Penicillium sp.<br>Phoene clematidis (Fuckel) S. Hughes<br>*Phoene clematidis (Fuckel) S. Hughes<br>*Phoene sp.<br>Sportdesmin sp.<br>*Trichocladium achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. achrasporum (Meyers & R.T. Moore) M. Dixon   | -<br>1<br>3<br>3<br>1<br>10<br>-<br>2<br>1<br>17<br>24<br>1<br>12<br>4   | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>-<br>-<br>-<br>2<br>0<br>3  | 22<br>15<br>-<br>-<br>-<br>1<br>-<br>-<br>1<br>-<br>-<br>1<br>-<br>-<br>1  |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Peniciality prolifica Anastasiou<br>*Phaeoisaria clematidis (Fuckel) S. Hughes<br>*Phoma sp.<br>Sporidesmium sp.<br>Frichocladium achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. alopallonellum (Meyers & R.T. Moore) M. Dixon<br>T. alopallonellum (Meyers & R.T. Moyers) Kohlm., & Voklm  | -<br>1<br>3<br>1<br>10<br>-<br>2<br>1<br>17<br>24<br>1<br>12<br>4<br>4   | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>-<br>-<br>-<br>20<br>3  | 22<br>15<br>-<br>-<br>-<br>1<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-   |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putrodinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Penicolia prolifica Anastasiou<br>*Phaeoisaria clematidis (Fuckel) S. Hughes<br>*Phone sp.<br>Spritsentidium achrosporum (Meyers & R.T. Moore) M. Dixon<br>T. achrosporum (Meyers & R.T. Moore) M. Dixon<br>T. achrosporum (Meyers & R.T. Moyers) Kohlm. & Voklm<br>Kohlm.  | -<br>1<br>3<br>1<br>10<br>-<br>2<br>1<br>17<br>24<br>1<br>12<br>4<br>4<br>4  | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>2<br>-<br>-<br>20<br>3<br>-   | 22<br>15<br>-<br>-<br>-<br>1<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-   |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Peniciillium sp.<br>Pericoil prolifica Anastasiou<br>*Phaeoissaria clematidis (Fuckel) S. Hughes<br>*Phoma sp.<br>Sportdesmium sp.<br>Sportdesmium sp.<br>*Trichocladium achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. alopallonellum (Meyers & R.T. Moore) M. Dixon<br>T. alopallonellum (Meyers & R.T. Meyers) Kohlm. & Voklm<br>Kohlm.  | -<br>1<br>3<br>1<br>10<br>-<br>2<br>1<br>17<br>24<br>1<br>12<br>4<br>4<br>4<br>4   | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>-<br>-<br>-<br>20<br>3<br>-<br>7  | 22<br>15<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-   |  |
| Dactylella sp.<br>Diplociadiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putrodinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Periocial prolifica Anastasiou<br>*Phaeoisaria clematidis (Fuckel) S. Hughes<br>*Phoma sp.<br>Sporidesmium sp.<br>Sporidesmium sp.<br>*Trichocladium achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. alopallouellum (Meyers & R.T. Moore) M. Dixon<br>T. alopallouellum (Meyers & R.T. Moore) M. Dixon<br>T. alopallouellum (Meyers & R.T. Moyers) & Nohlm. & VoklmKohlm   | -<br>1<br>3<br>3<br>1<br>10<br>-<br>2<br>1<br>1<br>24<br>1<br>12<br>4<br>4<br>4  | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>-<br>-<br>-<br>2<br>0<br>3<br>-<br>7<br>2   | 22<br>15<br>-<br>-<br>-<br>1<br>-<br>-<br>1<br>-<br>-<br>1<br>-<br>-<br>1<br>-<br>1  |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putrednits (Wallr.) S. Hughes<br>Monodictys sp.<br>Periodilium sp.<br>Princeilium sp.<br>*Phaeoiseria clematidis (Fuckel) S. Hughes<br>*Phona sp.<br>Sportdesmium sp.<br>*Trichocladium achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. achrasporum (Meyers & R.T. Moyers) Kohlm. & Voklm<br>Kohlm.<br>*T. linderi J.L. Crane & Shearer<br>Trichocladium sp.  | -<br>3<br>3<br>1<br>10<br>-<br>2<br>1<br>17<br>24<br>1<br>12<br>4<br>4<br>4<br>4<br>2<br>-<br>3  | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>2<br>2<br>-<br>-<br>20<br>3<br>-<br>-<br>20<br>3<br>-<br>-<br>7<br>2  | 22<br>15<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-   |  |
| Dactylella sp.<br>Diplociadiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Penicicilium sp.<br>Pericoili prolifica Anastasiou<br>*Phaeoisaria clematidis (Fuckel) S. Hughes<br>*Phoma sp.<br>Sporidesmium sp.<br>*Trichocladium achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. alopallouellum (Meyers & R.T. Moore) M. Dixon<br>T. alopallonellum (Meyers & R.T. Moyers) Kohlm. & Voklm<br>Kohlm.<br>*T. linderi J.L. Crane & Shearer<br>Trichocladium sp.<br>Verticillium sp.   | -<br>1<br>3<br>3<br>1<br>10<br>-<br>2<br>1<br>17<br>24<br>1<br>12<br>4<br>4<br>4<br>4<br>4<br>2<br>-<br>3<br>3                           | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>-<br>-<br>-<br>2<br>0<br>3<br>-<br>-<br>-<br>-<br>2<br>0<br>3<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-   | 22<br>15<br>-<br>-<br>-<br>1<br>-<br>-<br>-<br>1<br>-<br>-<br>-<br>1<br>-<br>-<br>-<br>1<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putrodinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Peniciality sp.<br>Penicality sp.<br>Periconia prolifica Anastasiou<br>*Prima sp.<br>Terichocladium achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. achrasporum (Meyers & R.T. Moyers) Kohlm. & Voklm<br>Kohlm.<br>*T. linderi J.L. Crane & Shearer<br>Trichocladium sp.<br>Yateriolium sp.   | -<br>1<br>3<br>1<br>10<br>-<br>2<br>1<br>17<br>24<br>1<br>12<br>4<br>4<br>4<br>4<br>2<br>-<br>3<br>13                                    | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>2<br>2<br>-<br>-<br>20<br>3<br>-<br>-<br>-<br>20<br>3<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-   | 22<br>15<br>-<br>-<br>-<br>1<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-   |  |
| Dactylella sp.<br>Diplociadiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Peniciality sp.<br>Periocial prolifica Anastasiou<br>*Phaeoisaria clematidis (Fuckel) S. Hughes<br>*Phoma sp.<br>Sporidesmium sp.<br>*Trichocladium achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. alopalloalium achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. alopalloanellum (Meyers & R.T. Meyers) Kohlm. & Voklm<br>Kohlm.<br>*T indori J.L. Crane & Shearer<br>Trichocladium sp.<br>Verticillium sp.<br>*Zalerion maritima (Linder) Anastasiou<br>*Z varia Anastasiou   | -<br>3<br>1<br>10<br>-<br>2<br>1<br>1<br>24<br>1<br>24<br>1<br>24<br>4<br>4<br>4<br>2<br>3<br>3<br>3<br>5                                | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>-<br>-<br>20<br>3<br>-<br>7<br>20<br>3<br>-<br>7<br>2<br>8<br>8   | 22<br>15<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-   |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Penicolia prolifica Anastasiou<br>*Phaeoisaria clematidis (Fuckel) S. Hughes<br>*Phoma sp. mp.<br>Marking and the state of the state of the state of the state<br>of the state of the state of the state of the state of the state<br>of the state of the state of the state of the state of the state<br>and state of the state of the state of the state of the state<br>for the state of the state of the state of the state of the state<br>for the state of the state of the state of the state of the state<br>of the state of the state of the state of the state of the state<br>of the state of the state of the state of the state of the state<br>for the state of the state of the state of the state of the state<br>for the state of the state<br>for the state of the | -<br>1<br>3<br>1<br>10<br>-<br>2<br>1<br>17<br>24<br>1<br>12<br>4<br>4<br>4<br>2<br>-<br>3<br>13<br>5<br>-                               | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>-<br>-<br>20<br>3<br>-<br>-<br>20<br>3<br>-<br>-<br>20<br>3<br>-<br>-<br>-<br>20<br>3<br>-<br>-<br>-<br>-<br>2<br>-<br>20<br>3<br>-<br>-<br>-<br>-<br>-<br>-<br>2<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 22<br>15<br>-<br>-<br>-<br>1<br>-<br>-<br>-<br>-<br>-<br>-<br>1<br>-<br>-<br>-<br>-<br>1<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putrednits (Wallr.) S. Hughes<br>Monodictys sp.<br>Periconia prelifica Anastasiou<br>Periconia prelifica Anastasiou<br>*Phaeoisaria clematidis (Fuckel) S. Hughes<br>*Phona sp.<br>Perichocladium achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. achrasporum (Meyers & R.T. Moyers) Kohlm. & Voklm<br>Kohlm.<br>*T. Inderi J. L. Crane & Shearer<br>Trichocladium sp.<br>*Zalerion maritima (Linder) Anastasiou<br>*Z. varia Anastasiou<br>Anamorph sp. 1   | -<br>3<br>1<br>10<br>-<br>2<br>1<br>17<br>24<br>1<br>12<br>4<br>4<br>4<br>4<br>2<br>-<br>3<br>13<br>5<br>-<br>-                          | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2  | 22<br>15<br>-<br>-<br>-<br>1<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-   |  |
| Dactylella sp.<br>Diplociadiella scalaroides G. Arnuad<br>Graphium sp.<br>Heliconyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Peniciality sp.<br>Periorial prolifica Anastasiou<br>*Phaeoisaria clematidis (Fuckel) S. Hughes<br>*Phoma sp.<br>Sporidesmium sp.<br>*Trichocladium achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. alopalloum achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. alopalloum linu (Meyers & R.T. Meyers) Kohlm. & Voklm<br>Kortichocladium achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. alopalloum achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. inderi J.L. Crane & Shearer<br>Trichocladium sp.<br>Yerticillium sp.<br>Yealerion maritima (Linder) Anastasiou<br>*Z. varia Anastasiou<br>Anamorph sp. 1<br>Anamorph sp. 2   | -<br>3<br>3<br>1<br>10<br>-<br>2<br>1<br>17<br>24<br>1<br>12<br>4<br>4<br>4<br>2<br>3<br>3<br>5<br>-<br>25                               | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>-<br>-<br>-<br>20<br>3<br>-<br>7<br>2<br>-<br>3<br>-<br>7<br>2<br>-<br>8<br>8<br>8<br>-<br>3<br>17  | 22<br>15<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-   |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Penicillium sp.<br>Penicillium sp.<br>Penicillium sp.<br>Phoeoidenia clematidis (Fuckel) S. Hughes<br>*Phoeoidenia sp.<br>*Tichocladium sp.<br>*Zalerion maritima (Linder) Anastasiou<br>*Zalerion maritima (Linder) Anastasiou<br>*Zalerion maritima (Linder) Anastasiou<br>*Zalerion hyp. 2<br>Total teleomorphs  | -<br>3<br>3<br>1<br>10<br>-<br>2<br>1<br>17<br>24<br>1<br>12<br>4<br>4<br>4<br>4<br>-<br>3<br>113<br>5<br>-<br>-<br>25<br>22             | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>-<br>-<br>-<br>20<br>3<br>-<br>-<br>-<br>20<br>3<br>-<br>-<br>-<br>8<br>8<br>-<br>-<br>8<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-   | 22<br>15<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-   |  |
| Dactylella sp.<br>Diplociadiella scalaroides G. Arnuad<br>Graphium sp.<br>Helicomyces roseus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys putredinis (Wallr.) S. Hughes<br>Peniccial prolifica Anastasiou<br>*Phaeoisaria clematidis (Fuckel) S. Hughes<br>*Phoma sp.<br>Sporidesmium sp.<br>*Princhocladium achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. adopalloulium achrasporum (Meyers & R.T. Meyers) Kohlm. & Voklm<br>Kohlm<br>*T. idnopalloum (Meyers & R.T. Meyers) Kohlm. & Voklm<br>Kohlm<br>*T. idnopalloum Sp.<br>Yealerion maritima (Linder) Anastasiou<br>*Zalerion maritima (Linder) Anastasiou<br>Anamoch sp. 2<br>Total teleomocphs<br>Total anamorphs<br>Total anamorphs   | -<br>1<br>3<br>1<br>10<br>-<br>2<br>1<br>17<br>24<br>1<br>12<br>4<br>4<br>4<br>2<br>-<br>3<br>13<br>5<br>-<br>-<br>25<br>22<br>247       | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>2<br>2<br>2<br>2<br>3<br>-<br>-<br>2<br>0<br>3<br>-<br>-<br>2<br>0<br>3<br>-<br>-<br>-<br>2<br>1<br>2<br>2<br>-<br>-<br>-<br>2<br>1<br>2<br>2<br>-<br>-<br>-<br>2<br>1<br>2<br>2<br>-<br>-<br>-<br>-                          | 22<br>15<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-   |  |
| Dactylella sp.<br>Diplocladiella scalaroides G. Arnuad<br>Graphium sp.<br>Heliconyces roxeus Link<br>*Monodictys putredinis (Wallr.) S. Hughes<br>Monodictys sp.<br>Peniciality orolifica Anastasiou<br>*Phaeoisaria clematidis (Fuckel) S. Hughes<br>*Phaeoisaria clematidis (Fuckel) S. Hughes<br>*Phaeoisaria clematidis (Fuckel) S. Hughes<br>*Phaeoisaria clematidis (Fuckel) S. Hughes<br>*Dichocladium achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. achrasporum (Meyers & R.T. Moore) M. Dixon<br>T. achrasporum (Meyers & R.T. Moyers) Kohlm. & Voklm<br>Kohlm.<br>*T. linderi J.L. Crane & Shearer<br>Trichocladium sp.<br>*Zalerion maritima (Linder) Anastasiou<br>*Z. varia Anastasiou<br>Anamorph sp. 1<br>Anamorph sp. 1<br>Anamorph sp. 1<br>Total teleomorphs<br>Total anamorphs   | -<br>1<br>3<br>1<br>10<br>-<br>2<br>1<br>17<br>24<br>1<br>24<br>1<br>12<br>4<br>4<br>4<br>-<br>3<br>13<br>5<br>-<br>25<br>222<br>47<br>2 | 27<br>6<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>- | 15<br>19<br>-<br>-<br>2<br>1<br>2<br>2<br>2<br>-<br>20<br>3<br>-<br>7<br>2<br>-<br>8<br>8<br>8<br>-<br>3<br>115<br>1528   | 22<br>15<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-<br>-   |  |



Fig.1. Per cent woody litter (natural and submerged) of Avicennia officinalis colonized by single and co-occurrence offilamentous fungi in Nethravathi mangrove.

*varia*) were common to natural and submerged wood in both seasons.

Shannon diversity of colonized fungi was the highest in natural wood in monsoon (4.542) followed by submerged wood in monsoon (4.351), submerged wood in summer (3.559) and natural wood in summer (3.291). The Pielou's equitability was highest in monsoon samples of submerged wood (0.870) followed by natural wood in monsoon (0.818), submerged wood in summer (0.798) and natural wood in summer 0.749).

**Co-occurrence**: Occurrence of single species on natural wood was ranged from 11-12%, while it was 10-27% in submerged wood (**Fig. 1**). Number of co-occurrences of species per wood was extended up to 11 and 8 species during monsoon in natural and submerged wood, respectively. In both wood types during summer, the maximum number co-occurrences of species per wood was only six species. In natural as well as submerged wood, co-occurrences of three species were the highest in both seasons. As seen in overall colonization (**Table 1**), the three species co-occurrences in natural wood were higher (teleomorphs, anamorphs and core-group species) than submerged wood, so also during monsoon than summer (**Table 2**).

On comparison of co-occurrences (%) of 20 core-group species, dominance was seen in natural than in submerged wood, so also during monsoon than summer (**Table 3**). Co-occurrence of core-group fungi was ranged from 3% (*Aigialus mangroveis* in submerged wood during summer) to 72% (*Lignincola laevis* in natural wood during summer). Single species occurrence of core-group fungi on wood was as low as 0% (in many species) and raised up to 12% (*Lignincola laevis* in natural wood during summer).

 Table 2.
 Colonization frequency (%) three species co-occurrence of fungi on Avicennia officinalis woody litter (-, not occurred).

| Taxon  | Natura  | l wood | Immersed wood |        |  |
|--|---------|--------|---------------|--------|--|
|  | Monsoon | Summer | Monsoon       | Summer |  |
| Ascomycetes  |         |        |               |        |  |
| Aigialus mangroveis Borse                          | -       | -      | 2             | 1      |  |
| Anintodera mangrovei K.D. Hyde                     | -       | -      | 1             | -      |  |
| Anintodera sp                                      | _       | _      | 2             | 4      |  |
| Camornalla rhizonhoraa Kohlm                       | 1       |        | 2             | -      |  |
| Under som heine singering stade Shaaran & LL Crone | 1       | 1      | 1             | -      |  |
| Halosarpheia cincinnatula Sheater & J.L. Crane     | -       | 1      | 1             | 0      |  |
| H. Jibrosa Kollilli. & E. Kollilli.                | 4       |        | -             | - 11   |  |
| Lightheoid idevis fiolitik                         | 4       | 24     | 12            | 11     |  |
| L. Iongirosiris (Chob & J. w. Chob) Kohili.        | -       | -      | 1             | -      |  |
| L. Hopica Romm.                                    | 2       | -      | -             | -      |  |
| Luiworinia granaispora Nicyels                     | -       | 2      | 12            | 22     |  |
| Luiworinia sp.                                     | -       | 3      | 15            | 25     |  |
| Passeriniella mangrovel G.L. Maria & K.K. Sridnar  | 14      | -      | 2             | 1      |  |
| Savoryella lignicola E.B.G. Jones & K.A. Eaton     | 5       | 4      | 2             | 2      |  |
| S. longispord E.B.G. Jones & K.D. Hyde             | 2       | 3      | -             | -      |  |
| S. paucispora (Cribb & J. W. Cribb) J. Koch        | 0       | 11     | 0             | 2      |  |
| Sporoschima sp.                                    | -       | -      | 2             | -      |  |
| Tirispora mandoviana V.V. Sarma & K.D. Hyde        | -       | -      | -             | 2      |  |
| <i>Tirispora</i> sp.                               | 4       | 11     | 13            | 18     |  |
| Verruculina enalia (Kohlm.) Kohlm. & Volkm. Kohlm. | -       | -      | 4             | 6      |  |
| Ascomycete sp. 1                                   | -7      | -      | 1             | -      |  |
| Ascomycete sp. 2                                   | 1       | -      | -             | -      |  |
| Ascomycete sp. 3                                   | -       | 1      | -             | -      |  |
| Ascomycete sp. 4                                   | -       | 5      | -             | -      |  |
| Anamorphs  |         |        |               |        |  |
| Acremonium sp.                                     | 2       | -      | -             | -      |  |
| Alternaria sp. 1                                   | 1       | -      | -             | -      |  |
| Cirrenalia pygmea Kohlm.                           | -       | 7      | 7             | 11     |  |
| C. tropicalis Kohlm.                               | -       | 2      | 7             | 8      |  |
| Diplocladiella scalaroides G. Arnuad               | 2       | -      | -             | -      |  |
| Graphium sp.                                       | 2       | -      | -             | -      |  |
| Monodictys putredinis (Wallr.) S. Hughes           | 5       | -      | -             | -      |  |
| Penicillium sp.                                    | 1       | -      | -             | -      |  |
| Periconia prolifica Anastasiou                     | -       | -      | 1             | -      |  |
| Phaeoisaria clematidis (Fuckel) S. Hughes          | 1       | -      | -             | -      |  |
| Phoma sp.  | 8       | -      | -             | -      |  |
| Sporidesmium sp.                                   | 1       | -      | -             | -      |  |
| Trichocladium achrasporum (Meyers & R.T. Moore) M. | 5       | -      | 7             | 2      |  |
| Dixon  |         |        |               |        |  |
| T. alopallonellum (Meyers & R.T. Meyers) Kohlm. &  | 1       | -      | -             | -      |  |
| VoklmKohlm.  |         |        |               |        |  |
| T. linderi J.L. Crane & Shearer                    | 16      | 1      | 4             | 1      |  |
| Trichocladium sp.                                  | -       | -      | 1             | -      |  |
| Verticillium sp.                                   | 2       | -      | -             | -      |  |
| Zalerion maritima (Linder) Anastasiou              | 2       | -      | 2             | 2      |  |
| Z. varia Anastasiou                                | 1       | 12     | 2             | 1      |  |
| Anamorph sp. 1                                     | -       | -      | 1             | -      |  |
| Anamorph sp. 2                                     | -       | -      | 1             | -      |  |
| Total teleomorphs                                  | 11      | 10     | 15            | 12     |  |
| Total anamorphs                                    | 15      | 4      | 10            | 6      |  |
| Total species                                      | 26      | 14     | 25            | 18     |  |
| Total core-group species (≥10%)                    | 2       | 5      | 3             | 4      |  |

**Table 3.** Comparison frequency (%) of co-occurrence of core-group fungi on *Avicennia officinalis* woody litter (within bracket, % wood colonized by only this species; -, not occurred; \*, occurrence <10%: see Table 1).

| Taxon  | Natural wood |         | Submerged wood |        |
|--|--------------|---------|----------------|--------|
|  | Monsoon      | Summer  | Monsoon        | Summer |
| Ascomycetes  |              |         |                |        |
| Aigialus mangroveis Borse                                | -            | -       | *              | 3 (8)  |
| Lignincola laevis Höhnk                                  | 16(1)        | 72 (12) | 28 (1)         | 22 (0) |
| L. tropica Kohlm.  | 10 (0)       | -       | *              | -      |
| Lulworthia grandispora Meyers                            | *            | 36 (0)  | *              | *      |
| Lulworthia sp.   | -            | *       | 38 (2)         | 48 (6) |
| Passeriniella mangrovei G.L. Maria & K.R. Sridhar        | 32 (6)       | -       | *              | *      |
| Savoryella lignicola E.B.G. Jones & R.A, Eaton           | 14 (0)       | *       | *              | *      |
| S. paucispora (Cribb & J.W. Cribb) J. Koch               | 28 (0)       | 18 (0)  | 17 (0)         | *      |
| Tirispora sp.  | *            | 24 (0)  | 33(1)          | 36 (6) |
| Verruculina enalia (Kohlm.) Kohlm. & Volkm. Kohlm.       | *            | -       | 12 (2)         | 16 (3) |
| Ascomycete sp. 1   | 21 (0)       | -       | -              | -      |
| Anamorphs  |              |         |                |        |
| Cirrenalia pygmea Kohlm.                                 | -            | 26 (0)  | 15 (0)         | 22 (1) |
| C. tropicalis Kohlm.                                     | -            | *       | 19 (0)         | 14 (0) |
| Monodictys putredinis (Wallr.) S. Hughes                 | 9 (0)        | -       | *              | -      |
| Phaeoisaria clematidis (Fuckel) S. Hughes                | 12 (0)       | -       | -              | -      |
| Phoma sp.  | 21 (0)       | -       | -              | -      |
| Trichocladium achrasporum (Meyers & R.T. Moore) M. Dixon | 12 (0)       | *       | 20(0)          | *      |
| T. linderi J.L. Crane & Shearer                          | 38 (4)       | *       | *              | *      |
| Zalerion maritima (Linder) Anastasiou                    | 12 (0)       | -       | *              | *      |
| Z. varia Anastasiou                                      | *            | 27 (0)  | *              | *      |

A few examples of high co-occurrence in our study includes 11 fungi on one natural wood in monsoon (Halosarpheia fibosa, Lignincola laevis, Lulworthia grandispora, Monodictys putredinis, Phaeoisaria clematidis, Saagaromyces ratnagiriensis, Savoryella paucispora, S. lignicola, Trichocladium linderi, Verruculina enalia and Zalerion maritima); 10 fungi on one natural wood in monsoon (Ascomycete sp. 1, Kallichroma tethys, Lignincola tropica, Lulworthia grandispora, Monodictys putredinis, Periconia prolifica, Phaeoisaria clematidis, Savoryella paucispora, Trichocladium achrasporum and T. linderi); eight fungi on one natural wood in monsoon (Aniptodera mangrovei, Ascomycete sp. 1, Lignincola laevis, Lulworthia grandispora, Savoryella lignicola, S. paucispora, Trichocladium linderi and Zalerion maritima). In these three combinations (11, 10 and 8 fungi) L. grandispora, S. paucispora and T. linderi were common. Similarly, eight species co-occurrence in one submerged wood include Aniptodera sp., Halosarpheia cincinnatula, Lignincola tropica, Lulworthia sp., Penicillium sp., Savoryella lignicola, *Tirispora* sp., and *Trichocladium achrasporum*.

# DISCUSSION

**Colonization and diversity:** Colonization and diversity of marine fungi are influenced by various biotic (e.g. nature of wood, extent of decay and fungal competition) and abiotic (e.g. duration of submersion and water chemistry) factors, where substrata play a significant role (Nakagiri, 1993; Hyde and Lee, 1995; Jones, 2000). The present study revealed variations in the pattern of fungal colonization, diversity and co-occurrence on woody litter of *Avicennia officinalis* as seen in earlier studies on woody litter of *Rhizophora mucronata* (Sridhar and Maria, 2006; Sarma and Raghukumar, 2013). Compared to *Rhizophora* wood, the overall fungal richness in *Avicennia* wood in mangroves was lower (e.g. Sarma and Vittal, 2000; Maria and Sridhar, 2003, 2004; Sridhar and Maria, 2006; Sarma and Raghukumar, 2013).

The dominance of fungi on woody litter in mangroves is dependent on the mangrove ecosystem. For instance, fungi those are dominant on wood in mangroves of Gujarat, Maharashtra and Goa were different than those found in the mangroves of Karnataka and Kerala (Maria and Sridhar, 2002, 2003, 2004; Sridhar and Maria, 2006; Sarma and Raghukumar, 2013; Sridhar 2013). Although some studies are available on the colonization and diversity of fungi on woody litter in mangroves of the Indian coast, comparative accounts between east coast and west coast are lacking.

As seen in the present study, naturally submerged wood of *Rhizophora mucronata* possess higher fungal occurrence than artificially submerged wood in mangroves (Sridhar and Maria, 2006). The dominance of fungi on mangrove wood will be more during monsoon than summer (Maria and Sridhar, 2003; Sridhar and Maria, 2006). The terrestrial fungi were dominant in overall occurrence during monsoon season likely owing to lower salinity than summer season. Considerable number of terrestrial fungi was also seen during monsoon in three species co-occurrences (see **Table 2**). Terrestrial fungi were not the members of core-group in natural as well as submerged wood (see **Table 3**). However,

the role of terrestrial fungi in mangroves during monsoon season cannot be ignored as they depend on ruderal strategy for colonization on substrata and dissemination.

In our study, the Shannon diversity was highest in monsoon than summer irrespective of natural and submerged wood of *A. officinalis*. The results were also similar in natural and submerged wood of *R. mucronata* in Nethravathi mangrove (Sridhar and Maria, 2006). The diversity on five types of naturally submerged mangrove woody litter also represented the higher diversity during monsoon than summer season (Maria and Sridhar, 2003), so also on deliberately submerged *Avicennia officinalis* as well as *Rhizophora mucronata* woody litter in the Udyavara mangrove of southwestern India (Maria and Sridhar, 2004). The fungal diversity was also higher during the monsoon season than summer season on naturally occurring woody litter in Nethravathi and Udyavara mangroves (Ananda and Sridhar, 2004).

**Co-occurrence:** The co-occurrence of fungi on woody litter in our study was extended up to 11 species on Avicennia officinalis, but co-occurrence was only up to 5-9 species in Rhizophora mucronata (Sridhar and Maria, 2006; Sarma and Raghukumar, 2013). On natural R. mucronata wood, two species dominance was predominant (21-40%) (Sridhar and Maria, 2006; Sarma and Raghukumar, 2013). In submerged R. mucronata wood, during monsoon three species cooccurrence was the highest (22%), while during summer two species co-occurrence was the highest (38%) (Sridhar and Maria, 2006). The impact of saprophytic fungi on wood in mangroves is largely influenced by co-occurrences or consortia rather than single species occurrence on woody litter (Sridhar and Maria, 2006; Sarma and Raghukumar, 2013). However, Rimora mangrovei occurred singly in 100% of Rhizophora mucronata wood which deserves further study. Based on co-culturing, Tan et al. (1995) demonstrated that Lignincola laevis and Verruculina enalia are inhibitory to each other. Miller et al. (1985) considered Lulworthia sp. as an aggressive fungus against other fungi. All these fungi on Avicennia wood were core-group fungi in our study. However, Lignincola laevis dominated on natural as well as submerged wood in both seasons (monsoon and summer), while Lulworthia sp. and Verruculina enalia were dominant only on submerged wood. Their co-occurrence on wood ranged between 12 and 72%, while single species occurrence ranged from 2-12%, indicating that they cope up at various degrees with associated fungi during wood decomposition. Unlike dominance of Lignincola laevis on Avicennia officinalis wood (natural as well as submerged), Lulworthia grandispora was dominant on R. mucronata wood (Sridhar and Maria, 2006). Three core-group fungi (Lulworthia grandispora, Lignincola laevis and Verruculina enalia) coexisted with several fungi on woody litter in mangrove ecosystem (Ananda and Sridhar, 2004). Interestingly, V. enalia showed 100% co-occurrence on Rhizophora mucronata wood in Chorao mangroves of Goa, while Rimora mangrovei showed 100% single species occurrence (Sarma and Raghukumar, 2013).

In our study, *Cirrenalia tropicalis*, *Lulworthia* sp. and *Verruculina enalia* were dominant (or core-group) only on

submerged wood (see **Table 3**) which shows their capability to colonize freshly submerged wood successfully. Among 20 core-group fungi, 16 species (in natural/submerged wood and or monsoon/summer) did not occurred as single species which denotes that they support co-occurrences. However, *Aigialus mangroveis, Lulworthia* sp., *Verruculina enalia* showed single species occurrence on wood ranging from 2-8% and such independent occurrence needs further study.

### CONCLUSIONS

Saprophytic fungi are largely co-occurring to exploit the detritus resource. However, dominance of single species or a group of species or consortia cannot be ruled out. The present study forecasted that the extent of colonization of filamentous fungi on woody litter of Avicennia officinalis is next to Rhizophora mucronata. In addition to typical marine fungi, several terrestrial fungi colonized the woody litter as demonstrated by earlier studies. Aniptodera sp., Lignincola laevis, Lulworthia grandispora, Savorvella lignicola, S. paucispora, Tricladium linderi, Tirispora sp. and Zalerion varia were common to natural and submerged wood in monsoon and summer seasons. The diversity was higher during monsoon than summer irrespective of type of wood corroborating the earlier findings. The co-occurrence of fungi per wood was as high as 11 species during monsoon on naturally submerged wood. The three fungal co-occurrences was highest in natural as well as introduced wood during both seasons. Lignincola laevis co-occurred up to a maximum of 72% on natural wood during summer which reflects its compatibility or accommodating ability with associated fungi. Several questions seem to be pertinent on fungal cooccurrences or consortia on a substrate: I) Do they accommodate any fungi ? II) Are they tolerant/antagonistic/ selectively inhibitory to each other ? III) Can they function synergistically in resource exploitation? IV) Do they follow successional pattern to exploit the resource? Conventional studies as well as precise molecular approaches provide further insight on colonization and co-occurrences of filamentous fungi on woody litter in mangrove ecosystems.

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