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# A new host record for Dictyoarthrinium sacchari (J.A. Stev.) Damon

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#### ABSTRACT

While investigating the mycological diversity of Sagar district of Central India, an interesting litter hyphomycetous fungus was recorded and studied, which have its association with leaves of *Cymbopogon citrates* (DC.) Stapf. The symptomatology, macroscopic and microscopic features of the present collection are in conformity with the diagnostic characters of *Dictyoarthrinium sacchari* (J.A. Stev.) Damon. It is also noteworthy that this fungal species has never been earlier reported on this host. Hence this constitutes a new host record for *Dictyoarthrinium sacchari*.

Key words:- Hyphomycetous fungi, Dictyoarthrinium central India, leaf litter

# INTRODUCTION

The forest floor constitutes a heterogenous group of plant remnants such as coarse fine deadwood, fallen twigs, leaves, needles, etc. These remnants form the important part of dead organic matter (Didion et al., 2014). There is great diversity and dynamic aggregation of litter decomposing fungi occurring in association with the forest floor. These fungiplay important role in the structural stability, nutrient availability, productivity and other critical factors of ecosystem functioning (Alanbagi et al., 2019). The freshly fallen leaves form a favorable substrate for the growth and establishment of these fungi. The presence of fungal mycelia form the basis for different types of ecological interactions with bacteria and invertebrates (Boddy et al., 2007, Hardoim et al., 2015; Prakash et al., 2015). Fungal communities show a clear succession in response to the availability of different types of substrate to be decomposed (Fukasawa et al., 2009; Purahong et al., 2016; Voříšková and Baldrian, 2013). During the surveys conducted in Sagar district of Madhya Pradesh an interesting fungus was found growing in association with the dead leaves of Cymbopogon citratus (DC.) Stapf. Based on the phenotypic characters it was identified as Dictyoarthrinium sacchari (J.A. Stev.) Damon with Cymbopogon citratus as a new host record (Damon, 1953).

#### MATERIAL AND METHODS

Collection of infected leaves was carried out and meanwhile they were taken to the laboratory for slide preparation using lactophenol cotton blue as a mounting medium. For the detailed study of morphology, light microscopy was utilized. 400X magnification was taken in application in order to determine the micrometry of mycelium, conidiophores, and conidia (30 each). However, for purpose of determining exact measurement and detailed morphology of fungus Scanning Electron Microscope was used. Since it was a dried collection, sample was not given preparative treatment for SEM observation (Piñar et al., 2015). Instead sample was coated with thin layer of gold-palladium using Denton Vacuum and observed using Nova Nano SEM 450. The fungal specimen was submitted to Ajrekar Mycological Herbarium (AMH), Agharkar Research Institute Pune, India and another specimen deposited in mycological herbarium (RSM), Department of Botany, Dr. Harisingh Gour University Sagar M. P. India.

#### TAXONOMIC DETAILS

*Dictyoarthrinium sacchari* (J.A. Stev.) Damon, Notes on the hyphomycetous genera, *Spegazzinia* Sacc. and *Isthmospora* 

Stevens. Bulletin of the Torrey Botanical Club **80** (3): 155-165, 1953. Figs. 1-2

= *Tetracoccosporium sacchari* Stevenson Johnston, J. R. and Stevenson J. A., 1917. Sugar-cane fungi and diseases of Porto Rico. *Dept. Agric. Porto Rico* 1(4): 177-264.

Colony effused irregularly spreading on the substratum, black. Mycelium superficial forming a network on host

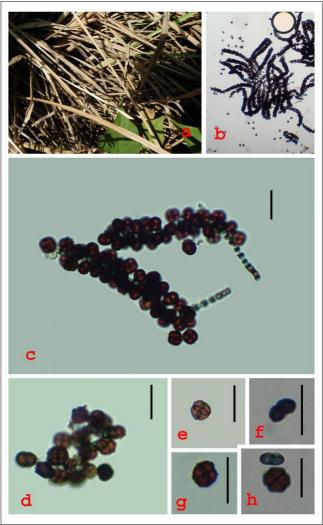
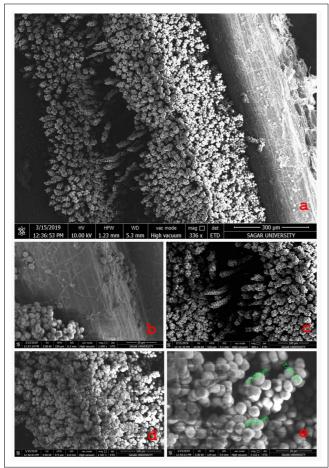


Fig. 1. *Dictyoarthrinium sacchari* a. Symptom. b-c. Conidia with conidiophores at 100X and 400X, respectively. d-h. Different view of conidia. Scale bar: c-h = 20µm



**Fig.2.** Scanning electron microscopic (SEM) images of *Dictyoarthrinium sacchari*: a. Fungus association with host. b. mycelium. c-e. Conidia in different magnification.

leaves, pale brown, hyphae  $1.7-2.9~\mu m$  thick. Conidiophores erect, cylindrical, aggregated, hyaline, smooth, up to 165~x  $4.5-5.0~\mu m$ . Conidiogenous cells forming dark bands on conidiophores. Conidia originate from lateral or apical part of conidiophores, dark brown, verrucose, thick walled, 4 celled, cruciately septate, initially 1-2 celled, oval, hyaline or pale brown, smooth,  $7.2-9.5~\mu m$  in size.

**Specimen examined:** Madhya Pradesh, Sagar, Dr. H. S. Gour Central University Sagar, Botanical Garden, on dead leaves of *Cymbopogon citratus*, Feb. 2018 leg. S. Bhardwaj, (AMH10046, RSM31).

### DISCUSSION

The present specimen resembles with the earlier description of *D. sacchari*. The review of literature revealed that *D. sacchari* is earlier reported in association with various hosts i.e. *Triticum* sp. (Nair and Tyagi, 1961), *Musa paradisiaca* (Matsushima, 1971), *Saccharum officinarum* (Stevenson, 1975), *Cosmos bipinnatus* (Srivastava and Gupta, 1981), *Lithachne pauciflora* (Mercado, 1984.), *Prunus amygdalus* (Gene *et al.*,1990), *Delonix elata* (Pandey and Rao ,1998), *Musa acuminata* (Photita *et al.*, 2003), *Neolitsea scrobiculata* (Saravanan and Vittal, 2007), *Persea mechrantha* (Saravanan

and Vittal, 2007) Saccharum spontaneum (Bhilabutra et al., 2010) and Pinus wallichiana (Prasher and Singh, 2015). The present report on Cymbopogon citratus is a new host record for D. sacchari.

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#### REFERENCES

- Alanbagi, R.A., Alshuwaili, F.E. and Stephenson, S.L. 2019. Fungi associated with forest floor litter in northwest Arkansas. *Current Research in Environmental & Applied Mycology* **9**(1): 25-35.
- Bhilabutra, W., McKenzie, E.H.C., Hyde, K.D. and Lumyong, S., 2010. Fungi on the grasses, *Thysanolaena latifolia* and *Saccharum spontaneum*, in northern Thailand. *Mycosphere* **1**(4): 301-314.
- Boddy, L., Frankland, J. and Van West, P. 2007. *Ecology of saprotrophic Basidiomycetes*. Academic Press. Elsevier, **28**: 365pp
- Damon, S. C., 1953. Notes on the hyphomycetous genera, Spegazzinia Sacc. and Isthmospora Stevens. Bulletin of the Torrey Botanical Club 80 (3): 155-165.
- Didion, M., Frey, B., Rogiers, N. and Thürig, E., 2014. Validating tree litter decomposition in the Yasso07 carbon model. *Ecological modelling* **291**: 58-68.
- Fukasawa, Y., Osono, T. and Takeda, H., 2009. Dynamics of physicochemical properties and occurrence of fungal fruit bodies during decomposition of coarse woody debris of *Fagus crenata*. *Journal of Forest Research* **14**(1): 20-29.
- Gene, J., Cano, J. and Guarro, J., 1990. Contribution to the study of the Spanish Hyphomycetes XI. *Revista Iberoamerican Micol*. 7: 31-33.
- Hardoim, P.R., Van Overbeek, L.S., Berg, G., Pirttilä, A.M., Compant, S., Campisano, A., Döring, M. and Sessitsch, A. 2015. The hidden world within plants: ecological and evolutionary considerations for defining functioning of microbial endophytes. *Microbiol. Mol. Biol. Rev.* **79**(3): 293-320.
- Johnston, J. R. and Stevenson J. A., 1917. Sugar-cane fungi and diseases of Porto Rico. Dept. Agric. Porto Rico 1(4): 177-264.
- Matsushima, T. 1971. *Microfungi of the Solomon Islands and Papua-New Guinea*. Published by the author. 78pp
- Mercado, A., 1984. Hifomicetes Demaciáceos de Sierra del Rosario, Cuba. Editorial Academia. 181pp

- Nair, N.G. and Tyagi, P.D., 1961, Notes on some hyphomycetes—I. In *Proceedings of the Indian Academy of Sciences*-Section B, Springer India. **54** (6): 269-275
- Pandey, A. and Rao, V.G., 1998. *A compendium of fungi on legumes from India*. Scientific Publishers (India) 188 pp
- Photita, W., Lumyong, P., McKenzie, E.H., Hyde, K.D. and Lumyong, S., 2003. Saprobic fungi on dead wild banana. *Mycotaxon* **85**: 345-356.
- Piñar, G., Sterflinger, K. and Pinzari, F., 2015. Unmasking the measles-like parchment discoloration: molecular and microanalytical approach. *Environmental microbiology* **17**(2): 427-443.
- Prakash, C.P., Thirumalai, E., Rajulu, M.G., Thirunavukkarasu, N. and Suryanarayanan, T.S., 2015. Ecology and diversity of leaf litter fungi during early-stage decomposition in a seasonally dry tropical forest. *Fungal ecology* 17: 103-113.
- Prasher, I.B. and Singh, G., 2015. New and interesting hyphomycetes from North-western Himalayas. *Kavaka* **44**: 83-86.

- Purahong, W., Wubet, T., Lentendu, G., Schloter, M., Pecyna, M.J., Kapturska, D., Hofrichter, M., Krüger, D. and Buscot, F., 2016. Life in leaf litter: novel insights into community dynamics of bacteria and fungi during litter decomposition. *Molecular Ecology*, **25**(16): 4059-4074.
- Saravanan, T. and Vittal, B.P.R. 2007. Some rare and interesting Hyphomycetes from Eastern Ghats in Tamil Nadu, India. *Kavaka* 35: 21-44.
- Srivastava, R.N. and Gupta, J.S. 1981. Seed mycoflora from Indian Seed lots of *Cosmos bipinatus* and their control. *Indian Phytopathol.* **34**(3), 383-385.
- Stevenson, J. A. 1975. Fungi of Puerto Rico and the American Virgin Islands. *Contr. Reed Herb.* **23**: 743.
- Voříšková, J. and Baldrian, P. 2013. Fungal community on decomposing leaf litter undergoes rapid successional changes. *The ISME Journal* 7(3): 477pp.